

The New England Journal of Medicine

©Copyright, 1996, by the Massachusetts Medical Society

Volume 334

FEBRUARY 8, 1996

Number 6

A CONTROLLED TRIAL OF TWO ACELLULAR VACCINES AND ONE WHOLE-CELL VACCINE AGAINST PERTUSSIS

DONATO GRECO, M.D., STEFANIA SALMASO, D.BIOL., PAOLA MASTRANTONIO, PH.D., MARINA GIULIANO, M.D., ALBERTO E. TOZZI, M.D., ALESSANDRA ANEMONA, D.STAT., MARTA L. CIOFI DEGLI ATTI, M.D., ANNA GIAMMANCO, PH.D., PIETRO PANEI, M.D., WILLIAM C. BLACKWELDER, PH.D., DAVID L. KLEIN, PH.D., STEVEN G.F. WASSILAK, M.D., AND THE PROGETTO PERTOSSE WORKING GROUP*

Abstract Background. Concern about both safety and efficacy has made the use of whole-cell pertussis vaccines controversial. In some European countries, including Italy, the rate of vaccination against pertussis is low.

Methods. We conducted a double-blind trial in Italy in which infants were randomly assigned to vaccination at two, four, and six months of age with an acellular pertussis vaccine together with diphtheria and tetanus toxoids (DTP); a DTP vaccine containing whole-cell pertussis (manufactured by Connaught Laboratories); or diphtheria and tetanus toxoids without pertussis (DT). The acellular DTP vaccine was either one containing filamentous hemagglutinin, pertactin, and pertussis toxin inactivated with formalin and glutaraldehyde (SmithKline Beecham) or one with filamentous hemagglutinin, pertactin, and genetically detoxified pertussis toxin (Chiron Biocine). Pertussis was defined as 21 days or more of paroxysmal cough, with infection confirmed by culture or serologic testing.

Results. The efficacy of each vaccine, given in three doses, against pertussis was determined for 14,751 chil-

dren over an average of 17 months, with cases included in the analysis if cough began 30 days or more after the completion of immunization. For both of the acellular DTP vaccines, the efficacy was 84 percent (95 percent confidence intervals, 76 to 89 percent for SmithKline DTP and 76 to 90 percent for Biocine DTP), whereas the efficacy of the whole-cell DTP vaccine was only 36 percent (95 percent confidence interval, 14 to 52 percent). The antibody responses were greater to the acellular vaccines than to the whole-cell vaccine. Local and systemic adverse events were significantly more frequent after the administration of the whole-cell vaccine. For the acellular vaccines, the frequency of adverse events was similar to that in the control (DT) group.

Conclusions. The two acellular DTP vaccines we studied were safe, immunogenic, and efficacious against pertussis, whereas the efficacy of the whole-cell DTP vaccine was unexpectedly low. (N Engl J Med 1996;334:341-8.)

©1996, Massachusetts Medical Society.

IN pediatrics the routine use of whole-cell vaccines against *Bordetella pertussis* has been a matter of continuous debate.¹⁻⁴ Acellular vaccines, consisting of purified proteins, have been in use for the primary immunization of two-year-old children in Japan since 1981.^{5,6} Two acellular vaccines were evaluated in a randomized clinical trial in Sweden,⁷ but the results left unanswered questions about the efficacy of the vaccines in infants, particularly in relation to that of whole-cell vaccines.

In Italy, vaccination of infants against diphtheria, tet-

anus, poliomyelitis, and hepatitis B is mandatory, but vaccination against pertussis is not. The rate of vaccination against pertussis varies greatly according to year of birth and geographic area.⁸ In 1991, the national average rate of vaccination against pertussis was estimated at 40 percent for children less than five years of age⁹; this low level of coverage can be attributed to the perception that there is an unacceptably high frequency of adverse events after the administration of the whole-cell vaccine.⁸ In 1992, we initiated the present randomized, double-blind, controlled clinical trial of three pertussis vaccines. The pertussis vaccines we studied were combined with diphtheria and tetanus toxoids and included a whole-cell vaccine currently used in the United States and two acellular vaccines, each containing inactive pertussis toxin, filamentous hemagglutinin, and pertactin. These proteins are involved in the pathogenesis of *B. pertussis* infection, and animal studies have suggested that they confer active immunity.^{10,11} Pertussis toxin has various biologic actions; because of its toxicity in animals, inactivation is required before it can be

From the Laboratory of Epidemiology and Biostatistics (D.G., S.S., A.E.T., A.A., M.L.C., P.P.) and the Laboratory of Bacteriology and Medical Mycology (P.M., M.G.), Istituto Superiore di Sanità, Rome; the Department of Hygiene and Microbiology, University of Palermo, Palermo, Italy (A.G.); and the National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda, Md. (W.C.B., D.L.K., S.G.F.W.). Address reprint requests to Dr. Greco at the Laboratory of Epidemiology and Biostatistics, Istituto Superiore di Sanità, viale Regina Elena 299, 00161 Rome, Italy.

Supported by a contract (N01-AI-25138) with the National Institute of Allergy and Infectious Diseases.

*The members of the Progetto Pertosse Working Group are listed in the Appendix.

used as an immunogen. The main objective of the trial was to examine the efficacy of each vaccine, given at two, four, and six months of age, in preventing laboratory-confirmed clinical pertussis.

METHODS

Participants

Infants were enrolled from September 1992 to September 1993 at 62 public health clinics operated by the National Health System, located in 4 of the 20 regions of Italy: Piemonte, Veneto, Friuli-Venezia Giulia, and Puglia. Nurses were hired and trained specifically to enroll and follow study children and to record information on standardized forms and in customized computerized data bases. The inclusion criteria for the study are shown in Table 1. The parents of each eligible newborn were invited to enter the trial; those who agreed gave written, informed consent. The study was approved by the Italian National Committee for Bioethics and the institutional review board of the U.S. National Institute of Allergy and Infectious Diseases (NIAID).

Vaccination

The composition of the study vaccines is shown in Table 2. The acellular diphtheria-tetanus-pertussis (DTP) vaccine manufactured by Chiron Biocine (Siena, Italy) contains genetically detoxified pertussis toxin,¹² filamentous hemagglutinin, and pertactin. The acellular DTP vaccine manufactured by SmithKline Beecham Biologicals (Rixensart, Belgium) contains pertussis toxin inactivated with formalin and glutaraldehyde, filamentous hemagglutinin, and pertactin. The heat-inactivated whole-cell DTP manufactured by Connaught Laboratories (Swiftwater, Pa.) belonged to a commercial lot licensed in the United States; this same lot was used concurrently in Sweden in a randomized

Table 2. Composition of the Vaccines Included in the Trial.

COMPONENT	SMITHKLINE ACELLULAR DTP VACCINE (LOT 116)	BIOCINE ACELLULAR DTP VACCINE (LOT 24 PFK/AH)	CONNAUGHT WHOLE-CELL DTP VACCINE (LOT 5299A)	BIOCINE DT VACCINE (LOT 141)
	<i>per 0.5-ml dose</i>			
Inactive pertussis toxin (μg)	25	5	Pertussis*	—
Filamentous hemagglutinin (μg)	25	2.5	Pertussis*	—
Pertactin (μg)	8	2.5	Pertussis*	—
Diphtheria toxoid (flocculation units)	25	25	6.65	25
Tetanus toxoid (flocculation units)	10	10	5	10
Aluminum-salt adjuvant	Aluminum hydroxide	Aluminum hydroxide	Aluminum phosphate	Aluminum hydroxide
Weight of ionic aluminum (mg)	0.5	0.35	0.15	0.7
Preservative	2-Phenoxyethanol	Thimerosal	Thimerosal	Thimerosal
Weight (mg)	2.5	0.05	0.05	0.05

*5.7 IU per dose by mouse intracerebral challenge test, as determined by the manufacturer.

clinical trial of two other acellular vaccines. A commercial diphtheria-tetanus (DT) preparation (Biocine) was used for the control group.

Three doses were administered when the infants were 6 to 12, 13 to 20, and 21 to 28 weeks of age, with a period of 4 to 12 weeks between successive doses. Criteria for discontinuing vaccination are included in Table 1. A booster dose of DT was given to all children six months after the third dose. The first two doses of the trial vaccine could be administered simultaneously with oral poliomyelitis vaccine and hepatitis B vaccine. The study vaccines were injected intramuscularly in the buttock or thigh.

Sample Size, Randomization, and Vaccine Masking

Assuming a true vaccine efficacy of 80 percent and a 5 percent incidence of laboratory-confirmed pertussis in unvaccinated children for the mean observation period, we calculated that a sample of 3300 children receiving each pertussis vaccine and 1100 receiving DT would provide an 85 percent probability that the lower limit of a two-sided 95 percent confidence interval for vaccine efficacy would be greater than 60 percent.¹³ Enrollment was augmented to compensate for attrition and potential overestimation of the incidence of pertussis. Ten sets of three doses each of vaccine were boxed together (three sets of each of the three DTP vaccines and one set of the DT vaccine, all in identical vials); the sets were consecutively numbered according to randomization lists provided by the NIAID. Infants were assigned to one of the four study groups at each clinic when they received the first dose of vaccine from the next available set of vials. Regular clinical personnel prepared and administered the vaccine. Neither parents nor investigators knew the infants' vaccine assignments.

Data Collection and Analysis

Efficacy

Nurses performed active surveillance by means of routine monthly telephone calls. Parents were instructed to note each of the child's episodes of coughing and report them to the study nurse. As soon as an illness characterized by cough lasting more than seven days was identified, samples of acute-phase serum and nasopharyngeal mucus were obtained; convalescent-phase serum was obtained six to eight weeks after the onset of cough. Parents recorded details of the illness and of treatment daily in a standardized diary; a nurse telephoned each week to review and record this information. The episode of coughing was considered over when the child had had no cough for two full days. Surveillance was conducted for coughing illnesses that began as late as December 31, 1994. Each child contributed time at risk for pertussis through the date of onset of laboratory-confirmed disease, the last date of contact with a study nurse, or the end of the surveillance period.

Vaccine efficacy was estimated for two periods, one beginning 30 days after the third dose of study vaccine and the other beginning im-

Table 1. Eligibility Criteria and Contraindications to Further Doses of Vaccine.

ELIGIBILITY CRITERIA
Medical
Age of 6–12 weeks
Weight >3rd percentile for age
No history of seizures or other central nervous system disease, including perinatal brain damage
No major congenital abnormalities, failure to thrive, or renal failure
No known or suspected immunologic deficit, including having a mother known to be positive for the human immunodeficiency virus
No prior illness compatible with pertussis
No prior vaccination against pertussis
Practical
Italian-speaking mother
Family accessible by telephone
Family planning to remain in the area for at least 12 months
CONTRAINDICATIONS TO FURTHER DOSES
Rectal temperature $\geq 40^{\circ}\text{C}$ within 48 hours
Persistent crying for ≥ 3 hours within 48 hours
Hypotonic, hyporesponsive episode or collapse within 48 hours
Generalized cyanosis within 48 hours
Anaphylaxis within 24 hours
Seizure, encephalitis, encephalopathy, or other serious central nervous system disease at any time

mediately after the first dose. A case of pertussis was defined as an illness with 21 days or more of paroxysmal cough and evidence of *B. pertussis* infection on culture or diagnostic serologic testing, as defined below. Vaccine efficacy after three doses was also estimated with the use of other clinical end points, according to the same laboratory criteria.

Immunogenicity

Paired capillary-blood samples were collected at each health center before the first dose of vaccine was given and one month after the third dose from a subsample of 10 percent of the children. Geometric mean titers of antibody to each pertussis antigen were measured before and after vaccination in each study group. A serologic response to each pertussis antigen was defined by a postvaccination titer at least four times higher than both the titer before vaccination and the minimal level of detection of the assay. Response to the diphtheria and tetanus toxoids was defined by a postvaccination antibody titer exceeding 0.01 IU per milliliter.

Safety

Parents recorded information on local and systemic symptoms in a standardized diary for the eight consecutive evenings after each vaccine dose was given. After the eighth day, nurses collected this information by telephone. If a serious illness occurred at any time, the study pediatricians verified the clinical history and reviewed all available documentation. Common side effects occurring during the two evenings after each dose were analyzed. Adverse events that were considered serious were anaphylaxis within 24 hours of vaccination; persistent crying (for 3 hours or more), a rectal temperature $\geq 40^{\circ}\text{C}$, hypotonic, hyporesponsive episodes, generalized cyanosis, or convulsions within 48 hours; and encephalopathy within 7 days.

Laboratory Procedures

Culture

Nasopharyngeal mucus was collected with an 8-French DeLee suction catheter (Sherwood Medical, St. Louis) and cultured on charcoal agar containing 10 percent defibrinated horse blood and 20 mg of cephalixin per liter (lot CM119, Unipath, Milan, Italy) in the regional laboratories. Isolates were identified by biochemical assay and by agglutination with specific antiserum (Murex Diagnostics, Dartford, England). All strains of *bordetella* were confirmed at the Istituto Superiore di Sanità in Rome.

Serologic Testing

Capillary blood was collected with Microtainer vials (Becton Dickinson, Rutherford, N.J.). All serologic assays were performed by personnel who had no knowledge of the infants' vaccine assignments or

the order of collection of the specimens; the samples were tested against pertussis antigens with reference serum calibrated against reference serum samples provided by the U.S. Food and Drug Administration (serum lot 3 or 4, Bethesda, Md.). Standardized enzyme-linked immunoassays (EIAs) were used to evaluate IgG and IgA antibodies to pertussis toxin and filamentous hemagglutinin and IgG antibodies to pertactin,¹⁴ with antigens provided by SmithKline Beecham. The reference-line method was used to calculate EIA units with standardized software (Unitcalc, Biosys inova, Stockholm, 1992). The minimal level of detection was set at 2 units per milliliter for IgG antibody to pertussis toxin and filamentous hemagglutinin, 3 units per milliliter for both IgG antibody to pertactin and IgA antibody to filamentous hemagglutinin, and 10 units per milliliter for IgA antibody to pertussis toxin. Any value below this minimum was recorded as half of the minimal value. The Chinese-hamster-ovary (CHO) assay of pertussis-toxin-neutralizing antibodies was performed only when sufficient serum was available.¹⁵ Neutralization titers were expressed as the reciprocal of the highest serum dilution that caused complete inhibition of typical clustering; the minimal level of detection was the first dilution tested (1:40), and undetectable values were recorded as 1:20.

For a serologic test to be considered positive, we required an increase in the antibody titer between acute-phase and convalescent-phase serum samples that was equivalent to one of the following: (1) an increase in the level of IgG or IgA antibody to pertussis toxin to twice the initial value; (2) an increase in the level of IgG or IgA antibody to filamentous hemagglutinin to twice the initial value, provided culture and the polymerase-chain-reaction (PCR) assay were negative for *B. parapertussis*; or (3) an increase in the level of pertussis-toxin-neutralizing antibodies to four times the initial value. A diagnostic doubling of the value on the EIA required the convalescent-phase antibody level to be at least four times the minimal level of detection.

Diphtheria antitoxin was assayed by toxin neutralization in Vero cells and tetanus antitoxin by a modified passive-hemagglutination assay^{16,17}; titers were converted to international units per milliliter with the use of an international reference serum.

B. parapertussis PCR

The PCR for the detection of *B. parapertussis* in aspirates involved the use of a specific insertion-sequence element IS1001.^{18,19}

Statistical Analysis

Vaccine efficacy was estimated as $1 - R$, where R is the ratio of the incidence of pertussis (ratio of cases to total person-time of follow-up) among the recipients of each DTP vaccine to the incidence among the control infants.²⁰ Confidence intervals were estimated by exact calculation,²¹ on the basis of the conditional binomial distribution of cases in one vaccine group and the total number of cases.²² Mean values for

Table 3. Confirmed Cases of Pertussis, Vaccine Efficacy, and Relative Risk, According to Vaccine Group and Number of Doses.*

VACCINE AND NO. OF DOSES	NO. OF CHILDREN	NO. OF PERSON-DAYS AT RISK	NO. OF CASES	INCIDENCE/100 PERSON-YEARS	VACCINE EFFICACY (95% CI)	RELATIVE RISK (95% CI)†
%						
SmithKline acellular DTP						
3 doses	4481	2,354,321	37	0.56	83.9 (75.8–89.4)	0.25 (0.17–0.36)
≥ 1 dose	4696	3,099,438	46	0.54	81.5 (73.1–87.4)	0.28 (0.20–0.39)
Biocine acellular DTP						
3 doses	4452	2,342,952	36	0.55	84.2 (76.2–89.7)	0.25 (0.17–0.36)
≥ 1 dose	4672	3,089,325	41	0.48	83.5 (75.6–88.9)	0.25 (0.17–0.36)
Connaught whole-cell DTP						
3 doses	4348	2,262,810	141	2.2	36.1 (14.2–52.1)	1.0
≥ 1 dose	4678	3,062,822	162	1.9	34.0 (12.8–49.8)	1.0
Biocine DT						
3 doses	1470	758,646	74	3.5	—	1.6 (1.2–2.1)
≥ 1 dose	1555	1,010,145	81	2.9	—	1.5 (1.1–2.0)

*Pertussis was defined clinically as 21 days or more of paroxysmal cough beginning 30 days or more after the third vaccine dose or immediately after the first dose. CI denotes confidence interval.

†Relative risks are expressed in relation to the incidence in the group given the whole-cell DTP vaccine.

Table 4. Vaccine Efficacy as Determined with Alternative Case Definitions, According to Vaccine Group.*

VACCINE	NO. OF PERSON-DAYS AT RISK	COUGH >7 DAYS		COUGH ≥14 DAYS		COUGH ≥21 DAYS		COUGH ≥30 DAYS		COUGH ≥60 DAYS		PAROXYSMAL COUGH ≥14 DAYS OR ANY COUGH ≥21 DAYS		PAROXYSMAL COUGH ≥14 DAYS	
		NO. OF CASES	VACCINE EFFICACY (95% CI)	NO. OF CASES	VACCINE EFFICACY (95% CI)	NO. OF CASES	VACCINE EFFICACY (95% CI)	NO. OF CASES	VACCINE EFFICACY (95% CI)	NO. OF CASES	VACCINE EFFICACY (95% CI)	NO. OF CASES	VACCINE EFFICACY (95% CI)	NO. OF CASES	VACCINE EFFICACY (95% CI)
		%		%		%		%		%		%		%	
SmithKline acellular DTP	2,354,321	84	70.6 (60.0–78.4)	78	72.7 (62.6–80.0)	58	79.0 (70.4–85.2)	40	84.1 (76.5–89.4)	15	90.3 (82.5–95.0)	69	75.6 (66.2–82.4)	55	78.4 (69.2–84.9)
Biocine acellular DTP	2,342,952	82	71.1 (60.7–78.8)	73	74.3 (64.7–81.4)	63	77.1 (68.0–83.7)	48	80.8 (72.2–86.9)	18	88.3 (79.7–93.6)	68	75.8 (66.5–82.6)	49	80.6 (72.1–86.7)
Connaught whole-cell DTP	2,262,810	211	23.1 (0.7–40.1)	202	26.4 (4.8–42.7)	194	26.9 (5.0–43.4)	173	28.4 (5.6–45.3)	96	35.6 (7.5–54.7)	197	27.4 (5.9–43.7)	167	31.7 (9.9–47.9)
Biocine DT	758,646	92	—	92	—	89	—	81	—	50	—	91	—	82	—

*Cases were included if cough began 30 days or more after the third vaccine dose. CI denotes confidence interval.

continuous variables were compared by means of the Kruskal–Wallis test. Differences between proportions were assessed by the chi-square test or Fisher's exact test. No adjustment of P values was made for multiple comparisons. The Wilcoxon rank-sum test was used to compare mean log antibody titers.

RESULTS

A total of 15,601 infants (49.6 percent girls and 50.4 percent boys), making up about 25 percent of the eligible newborns, received the first study dose at a mean age of 10.5 weeks; 15,101 received the second dose (mean age, 17.8 weeks); and 14,832 received the third (mean age, 24.9 weeks). In each vaccine group, 89 percent of the doses were injected into the buttocks. At the time the first two trial doses were given, 92 percent of each vaccine group also received hepatitis B vaccine and 94 percent received oral poliomyelitis vaccine.

Efficacy

Of the 15,601 enrolled children, 769 did not receive three doses of study vaccine. The reported reasons for the failure to administer the three-dose series were similar among the study groups, except for withdrawals due to side effects of the vaccine, which were more frequent after receipt of the whole-cell DTP vaccine; 135 children were withdrawn because of side effects after receiving the whole-cell DTP vaccine, 14 the SmithKline acellular DTP vaccine, 17 the Biocine acellular DTP vaccine, and 6 the DT vaccine. Of the 14,832 children who received three doses, 19 received doses of different products, 3 received partial doses, and 59 were excluded from further observation within 29 days after the administration of the third dose (13 because they had laboratory-confirmed illness and 46 who were lost to follow-up); therefore, 14,751 children (94.6 percent of those who were randomly assigned to groups) were included in the analysis of the efficacy of three doses of vaccine.

No significant differences were detected among these 14,751 children according to study group in terms of sex, age at vaccination, household size, number of children less than 13 years of age in the same household, or mean number of days of observation. The mean

length of follow-up for these children was 523 days (17.2 months), beginning 30 days after the third dose of vaccine. A total of 5147 episodes of cough lasting more than seven days were reported, and biologic specimens were collected for 4942 (96.0 percent) of these episodes, after a median of eight days of cough. Of 474 episodes of cough that were confirmed by laboratory testing to be associated with *B. pertussis*, 288 were defined as cases (with 21 days or more of paroxysmal cough) (Table 3). The average age at the onset of pertussis in these 288 cases was 551 days (18.2 months). The only complication noted after pertussis was a seizure in a recipient of the DT vaccine, who recovered without sequelae. The proportion of cases confirmed by culture was 73 percent for the whole-cell DTP vaccine, 82 percent for the DT vaccine, 76 percent for the SmithKline acellular DTP vaccine, and 67 percent for the Biocine acellular vaccine; the remaining cases were confirmed by serologic assays. Each of the two types of acellular vaccine was 84 percent efficacious after three doses, whereas the efficacy of the whole-cell DTP vaccine was 36 percent (Table 3). Estimates of vaccine efficacy based on only the 216 culture-confirmed cases were 85 percent for the SmithKline acellular DTP vaccine, 87 percent for the Biocine acellular vaccine, and 43 percent for the whole-cell DTP vaccine. Examination of alternative clinical criteria for confirmed *B. pertussis* infection indicates that the vaccine efficacy after three doses increased for illnesses with increasingly longer durations of cough, but it nonetheless remained low for the whole-cell DTP vaccine (Table 4).

For the entire group of 15,601 children randomly assigned to vaccine groups, biologic specimens were collected for 5152 episodes of cough over an average of 21.6 months of follow-up after the first dose. There were 531 episodes of cough laboratory-confirmed as associated with *B. pertussis*, of which 343 were cases with paroxysmal cough lasting 21 days or more (Table 3); for each vaccine, vaccine efficacy after the first dose was similar to that after three doses.

A total of 24 cases occurred from randomization



through the 29 days after the third dose (8 in recipients of the SmithKline acellular vaccine, 1 in a recipient of the Biocine acellular vaccine, 12 in recipients of the whole-cell DTP vaccine, and 3 in recipients of the DT vaccine) — an insufficient number to permit meaningful estimates of the incremental efficacy of each dose. Another analysis was based on cumulative periods from 30 days after the first dose to either 29 days after the third dose or 231 days of age, if fewer than three doses were received (this corresponded to 29 days after the maximal age at which the third dose could be given). In this period, 5 cases occurred in recipients of the SmithKline acellular vaccine (for an incidence of 0.39 per 100 person-years), 1 in a recipient of the Biocine acellular vaccine (0.08 per 100 person-years), 10 in recipients of the whole-cell DTP vaccine (0.79 per 100 person-years), and 2 in recipients of the DT vaccine (0.48 per 100 person-years). In pairwise comparisons, the incidence in recipients of the Biocine acellular DTP vaccine differed significantly from that in recipients of the whole-cell DTP vaccine (exact $P=0.006$, assuming the binomial distribution).²³

Immunogenicity

Serum specimens were obtained both before and after vaccination from 1572 children; 808 were tested by the CHO assay (Table 5). Antibody titers in the specimens obtained before immunization did not differ significantly among the groups. Each acellular vaccine elicited significantly higher titers of IgG and neutralizing antibody to pertussis toxin than did the whole-cell vaccine; the Biocine acellular DTP vaccine induced higher titers than the SmithKline acellular vaccine. In each group given an acellular DTP vaccine, the rate of serologic response on EIA was 94 percent or higher; the proportion

with a serologic response by CHO assay was significantly higher for the Biocine acellular vaccine than for the SmithKline product. For the whole-cell DTP vaccine, the proportion of children with a serologic response to pertussis toxin by EIA or by CHO assay was minimal. Geometric mean titers of IgG antibody to filamentous hemagglutinin and pertactin were much higher after vaccination with the acellular DTP vaccine than with the whole-cell vaccine; in recipients of the SmithKline acellular DTP vaccine, titers were significantly higher than in recipients of the Biocine acellular DTP vaccine. The proportions of children with a serologic response to filamentous hemagglutinin differed significantly among the vaccine groups. For the two acellular DTP vaccines, the serologic response to pertactin was more than 95 percent, whereas it was significantly lower after the administration of the whole-cell DTP vaccine.

Safety

Table 6 shows the incidence of adverse events within the first two days after vaccination. The common events we investigated were significantly more frequent in the group given the whole-cell DTP vaccine; overall, the frequency of these events in the two groups given acellular DTP vaccine was similar to that in DT-vaccine recipients. Swelling was more frequently reported after the administration of the acellular DTP vaccine than after the administration of the DT vaccine; it was somewhat more frequent in the children who received SmithKline acellular DTP vaccine than in those who received the Biocine acellular vaccine. Rectal temperatures $\geq 38^{\circ}\text{C}$ were infrequent after vaccination with either of the acellular DTP vaccines but occurred significantly more often than in the DT-vaccine recipients; such temperatures were more frequent in recipients of the SmithKline

Table 5. Geometric Mean Titers of Antibodies to Indicated Antigens and Rate of Serologic Response, According to Vaccine Group.*

ASSAY	SMITHKLINE ACELLULAR DTP VACCINE		BIOCINE ACELLULAR DTP VACCINE		CONNAUGHT WHOLE-CELL DTP VACCINE		BIOCINE DT VACCINE	
	GMT (95% CI)	RESPONSE (%)	GMT (95% CI)	RESPONSE (%)	GMT (95% CI)	RESPONSE (%)	GMT (95% CI)	RESPONSE (%)
EIA for IgG to pertussis toxin (units/ml)	51.3 (47.9–54.9)†	94.5†	94.4 (88.8–100.3)†‡	96.7†	1.2 (1.1–1.3)	4.2	1.0 (1.0–1.1)	—
EIA for IgG to filamentous hemagglutinin (units/ml)	147.0 (138.3–156.2)†§	85.1†¶	52.6 (49.1–56.3)†	60.5†¶	5.2 (4.7–5.8)	13.1¶	1.5 (1.3–1.6)	—
EIA for IgG to pertactin (units/ml)	274.2 (253.6–296.7)†§	96.6†	136.6 (127.0–146.8)†	95.9†	9.9 (8.6–11.3)	37.9	1.6 (1.6–1.7)	—
Pertussis-toxin neutralization by CHO, reciprocal of end dilution	230.0 (203.7–259.7)†	67.8†	787.6 (718.2–863.5)†‡	93.6†‡	23.0 (21.4–24.6)	1.7	22.0 (20.2–23.9)	—
Diphtheria		96.6**		98.8††		92.9		98.8‡‡
Tetanus		99.8		100		99.1		100

*GMT denotes the geometric mean titer after vaccination, CI confidence interval, EIA enzyme-linked immunoassay, and CHO Chinese-hamster-ovary assay.

† $P<0.001$ for the comparisons with the whole-cell DTP vaccine and the DT vaccine.

‡ $P<0.001$ for the comparison with the SmithKline acellular DTP vaccine.

§ $P<0.001$ for the comparison with the Biocine acellular DTP vaccine.

¶ $P<0.001$ for each pairwise comparison of DTP-vaccine groups.

||Geometric means for diphtheria and tetanus were not calculated because determinations were not made to the last dilution.

** $P=0.015$ for the comparison with the whole-cell DTP vaccine.

†† $P=0.046$ for the comparison with the SmithKline acellular DTP vaccine; $P<0.001$ for the comparison with the whole-cell DTP vaccine.

‡‡ $P=0.009$ for the comparison with the whole-cell DTP vaccine.

Table 6. Adverse Events within Two Days of Vaccination, According to Vaccine Group.*

EVENT	SMITHKLINE ACELLULAR DTP VACCINE (13,761 DOSES)	BIOCINE ACELLULAR DTP VACCINE (13,713 DOSES)	CONNAUGHT WHOLE-CELL DTP VACCINE (13,520 DOSES)	BIOCINE DT VACCINE† (4540 DOSES)
	<i>no. of events (rate/1000 doses)</i>			
Local swelling	1236 (90)‡‡	965 (70)§	3512 (260)¶	279 (61)
Local tenderness	628 (46)	625 (46)	4011 (297)¶	202 (45)
Irritability	4132 (300)	4153 (303)	6736 (499)¶	1351 (298)
Fever				
Rectal temperature $\geq 38.0^{\circ}\text{C}$	983 (72)‡‡	584 (43)¶	5425 (405)¶	151 (34)
Rectal temperature $\geq 40.0^{\circ}\text{C}$	5 (0.36)	4 (0.29)	32 (2.4)**	2 (0.44)
Persistent crying for ≥ 3 hours	6 (0.44)	9 (0.66)	54 (4.0)¶	—
Hypotonic, hyporesponsive episodes	—	1 (0.07)	9 (0.67)‡‡	2 (0.44)
Generalized cyanosis	—	—	2 (0.15)	—
Seizures	1 (0.07)	—	3 (0.22)	—

*Local events, irritability, and rectal temperature $\geq 38^{\circ}\text{C}$ are included if they occurred within two evenings after the administration of a vaccine dose; other events, within 48 hours of vaccination. The denominators used in calculating event rates may differ because of missing data.

‡ $P < 0.001$ for the comparison with the Biocine acellular DTP vaccine.

§ $P < 0.001$ for the comparison with the DT vaccine.

¶ $P = 0.038$ for the comparison with the DT vaccine.

‡‡ $P < 0.001$ for the comparisons with each of the other vaccine groups.

¶¶ $P = 0.005$ for the comparison with the DT vaccine.

** $P < 0.001$ for the comparison with the SmithKline acellular DTP vaccine and with the Biocine acellular DTP vaccine; $P = 0.010$ for the comparison with the DT vaccine.

‡‡‡ $P = 0.010$ for the comparison with the Biocine acellular DTP vaccine; $P = 0.002$ for the comparison with the Smith-Kline acellular DTP vaccine.

vaccine than in recipients of the Biocine vaccine. In all but the recipients of the whole-cell DTP vaccine, the frequency of temperatures $\geq 38^{\circ}\text{C}$ and local symptoms increased with each dose in the series.

Rectal temperature $\geq 40^{\circ}\text{C}$, crying for three hours or more, and hypotonic, hyporesponsive episodes were rarely reported after the administration of the DT vaccine or the acellular DTP vaccines; the frequency of these symptoms after the administration of the whole-cell DTP vaccine was greater than in all the other groups. Although the frequency of temperatures $\geq 40^{\circ}\text{C}$ increased with each additional dose, persistent crying for three hours or more and hypotonic, hyporesponsive episodes were more frequently reported after the first dose. No episodes of anaphylaxis or encephalopathy were observed. All children who had severe events within 48 hours of vaccination recovered without sequelae.

DISCUSSION

In this study, primary vaccination in infancy with either of two types of three-component acellular DTP vaccine was found to be highly effective in preventing clinical, laboratory-confirmed pertussis. Virtually all recipients of the DT vaccine who had confirmed disease had cough for 21 days or more (89 of 92 children), whereas the corresponding proportions in the recipients of the acellular DTP vaccines were lower (Smith-Kline, 58 of 84 children; Biocine, 63 of 82 children). Inclusion of a randomly assigned control group given only the DT vaccine provided precise estimates of absolute

vaccine efficacy, without which estimates of efficacy in relation to that of the whole-cell vaccine could have been uninformative. The use of a U.S.-licensed whole-cell vaccine that was also included in the clinical trial conducted by the Swedish Institute for Infectious Disease Control²⁴ provided a solid basis for interpreting the results of both trials.

In the Swedish trial, the observed efficacy of the same whole-cell DTP vaccine was 48 percent (95 percent confidence interval, 37 to 58 percent); a five-component vaccine containing 10 μg of pertussis toxin inactivated with glutaraldehyde, 5 μg each of filamentous hemagglutinin and fimbrial antigens (serotypes 2 and 3), and 3 μg of pertactin was highly efficacious (85 percent), whereas a two-component vaccine containing 25 μg each of filamentous hemagglutinin and pertussis toxin inactivated with formalin and glutaraldehyde had inferior efficacy (59 percent).²⁴ In another placebo-controlled, randomized clinical trial in Sweden, 40 μg of peroxide-inactivated pertussis toxin administered at 3, 5, and 12 months of age was found to be 71 percent effective (95 percent confidence interval, 63 to 78 percent) in preventing laboratory-confirmed disease characterized by 21 days or more of paroxysmal coughing.²⁵

The results of our trial are supported by a case-contact study of secondary transmission in households, in which the same three-component acellular DTP vaccine (SmithKline) was administered at two, four, and six months of age, with an estimated efficacy of 89 percent (95 percent confidence interval, 77 to 95 percent).²⁶ Results of other studies of acellular vaccines are expected shortly.^{27,28} Different vaccine preparations, vaccination schedules, study designs, or determinations of outcome variables can limit the comparability of study results, however.

The potential for bias exists in the laboratory confirmation of cases, since confirmation may be more limited in vaccinated children.²⁹ However, with clinical pertussis defined as 21 days or more of paroxysmal coughing, there appears to be no substantial bias; point estimates of vaccine efficacy were minimally altered when we considered only culture-confirmed cases.

In the present trial, the efficacy of the two acellular DTP vaccines was similar despite differences in the methods of inactivating pertussis toxin and the amounts of the various antigens included in the vaccines. Although antibody responses to each acellular vaccine were high, differences in geometric mean antibody titers were observed between the two acellular vaccines. Antibody

responses to the other antigens reflected the quantity of vaccine per dose, but titers of antibody to pertussis toxin did not correspond with the weight of antigens in each dose; this is consistent with other reports that the genetically inactivated pertussis toxin induces a stronger antibody response than chemically detoxified toxin.³⁰ The weak antibody response to pertussis toxin and filamentous hemagglutinin induced by the Connaught whole-cell DTP vaccine has been reported in other studies.^{31,32}

The level of protection conferred by the whole-cell vaccine was lower than we anticipated. Caution must be used in interpreting differences in serologic responses to vaccines, however, since serologic responses to specific pertussis antigens have not been shown to be correlated with clinical protection.⁷ Despite this fact, and although responses were measured only to the antigens contained in the acellular DTP vaccines, these results suggest that the protective effectiveness of whole-cell vaccines should be questioned if their immunogenicity is low.^{33,34} Studies of whole-cell pertussis vaccines in general report a higher level of protective efficacy than we found, albeit with differences in methods and the possible influence of other factors, such as the administration of booster doses.^{28,35,36}

The frequencies of common and uncommon adverse events after the administration of the whole-cell DTP vaccine were similar to the rates previously reported for whole-cell vaccines and much higher than those observed in recipients of the acellular vaccines.³⁷⁻³⁹ There were only minimal differences between the two acellular DTP vaccines and the DT vaccine in this respect. Because of the high observed efficacy and the improved safety profile of the acellular DTP vaccines, their use for the immunization of infants appears highly preferable to the continued use of the whole-cell vaccines.

We are indebted to V. Rafti and his staff for their full engagement in the administrative management of the trial; to M. Kanieff for valuable assistance throughout the study; to the staff of the NIAID for technical support; to the members of the Steering Committee and the Data Safety Monitoring Committee, in particular to Professor G.M. Fara, who acted as safety officer; and to the staff members of the participating local health units, who actively contributed to the conduct of the study.

APPENDIX

The members of the Progetto Pertosse Working Group in Italy were as follows: *Laboratory of Bacteriology and Medical Mycology* — P. Stefanelli, M. Bottone, and T. Sofia; *Laboratory of Epidemiology and Biostatistics* — S. Luzi, G. Bellomi, F. Cobianchi, G. Cangarella, and F. Meduri; *Laboratory of Immunology, Istituto Superiore di Sanità, Rome* — G. Scuderi; *Department of Hygiene and Microbiology, University of Palermo, Palermo* — A. Chiarini, M. Maggio, S. Taormina, and M. Genovese; *Piemonte region* — A. Moiraghi, A. Barale, S. Di Tommaso, S. Malaspina, and E. Vasile; *Veneto region* — P. Ferraro, P. Dal Lago, L. De Marzi, L. Robino, and E. Giraldo; *Friuli-Venezia Giulia region* — N. Coppola, P. Materassi, G. Tarabini Castellani, and F. Basso; and *Puglia region* — S. Barbuti, M. Quarto, P. Lopalco, P. D'Orazio, and A. Sanguedolce.

REFERENCES

1. Kanai K. Japan's experience in pertussis epidemiology and vaccination in the past thirty years. *Jpn J Med Sci Biol* 1980;33:107-43.

2. Miller DL, Alderslade R, Ross EM. Whooping cough and whooping cough vaccine: the risks and benefits debate. *Epidemiol Rev* 1982;4:1-24.
3. Romanus V, Jonsell R, Bergquist S-O. Pertussis in Sweden after the cessation of general immunization in 1979. *Pediatr Infect Dis J* 1987;6:364-71.
4. Howson CP, Howe CJ, Fineberg HV. Adverse effects of pertussis and rubella vaccines: a report of the Committee to Review the Adverse Consequences of Pertussis and Rubella Vaccines. Washington, D.C.: National Academy Press, 1991.
5. Kimura M, Kuno-Sakai H. Current epidemiology of pertussis in Japan. *Pediatr Infect Dis J* 1990;9:705-9.
6. Mortimer EA Jr, Kimura M, Cherry JD, et al. Protective efficacy of the Takeda acellular pertussis vaccine combined with diphtheria and tetanus toxoids following household exposure of Japanese children. *Am J Dis Child* 1990;144:899-904.
7. Ad Hoc Group for the Study of Pertussis Vaccines. Placebo-controlled trial of two acellular pertussis vaccines in Sweden — protective efficacy and adverse events. *Lancet* 1988;1:955-60. [Erratum, *Lancet* 1988;1:1238.]
8. The Italian Vaccine Coverage Survey Working Group. Childhood vaccination coverage in Italy: results of a seven-region survey. *Bull World Health Organ* 1994;72:885-95.
9. Binkin NJ, Salmaso S, Tozzi AE, Scuderi G, Greco D. Epidemiology of pertussis in a developed country with low vaccination coverage: the Italian experience. *Pediatr Infect Dis J* 1992;11:653-61.
10. Oda M, Cowell JL, Burstyn DG, Manclark CR. Protective activities of the filamentous hemagglutinin and the lymphocytosis-promoting factor of *Bordetella pertussis* in mice. *J Infect Dis* 1984;150:823-33.
11. Shahin RD, Brennan MJ, Li ZM, Meade BD, Manclark CR. Characterization of the protective capacity and immunogenicity of the 69-kD outer membrane protein of *Bordetella pertussis*. *J Exp Med* 1990;171:63-73.
12. Pizza M, Covacci A, Bartoloni A, et al. Mutants of pertussis toxin suitable for vaccine development. *Science* 1989;246:497-500.
13. Blackwelder WC. Sample size and power for prospective analysis of relative risk. *Stat Med* 1993;12:691-8.
14. Manclark CR, Meade BD, Burstyn DG. Serological response to *Bordetella pertussis*. In: Rose NR, Friedman H, Fahey JL, eds. *Manual of clinical laboratory immunology*. 3rd ed. Washington, D.C.: American Society for Microbiology, 1986:388-94.
15. Gillenius P, Jäättmäa E, Askelöf P, Granström M, Tiru M. The standardization of an assay for pertussis toxin and antitoxin in microplate culture of Chinese hamster ovary cells. *J Biol Stand* 1985;13:61-6.
16. Miyamura K, Tajiri E, Ito A, Murata R, Kono R. Micro cell culture method for determination of diphtheria toxin and antitoxin titres using VERO cells. II. Comparison with the rabbit skin method and practical application for seroepidemiological studies. *J Biol Stand* 1974;2:203-9.
17. Pitzurra M, Bistoni F, Pitzurra L, Marconi B. Use of turkey red blood cells in the passive haemagglutination test for studying tetanus immunity. *Bull World Health Organ* 1983;61:331-8.
18. van der Zee A, Agterberg C, van Agterveld M, Peeters M, Mooi FR. Characterization of IS1001, an insertion sequence element of *Bordetella parapertussis*. *J Bacteriol* 1993;175:141-7.
19. van der Zee A, Agterberg C, Peeters M, Schellekens J, Mooi FR. Polymerase chain reaction assay for pertussis: simultaneous detection and discrimination of *Bordetella pertussis* and *Bordetella parapertussis*. *J Clin Microbiol* 1993;31:2134-40.
20. Breslow NE, Day NE. *Statistical methods in cancer research. Vol. 2. The design and analysis of cohort studies*. Lyon, France: International Agency for Research on Cancer, 1987. (IARC scientific publications no. 82.)
21. Clopper CJ, Pearson ES. The use of confidence or fiducial limits illustrated in the case of the binomial. *Biometrika* 1934;26:404-13.
22. Johnson NL, Kotz S. *Discrete distributions*. Boston: Houghton Mifflin, 1969.
23. Kleinbaum DG, Kupper LL, Morgenstern H. *Epidemiologic research: principles and quantitative methods*. Belmont, Calif.: Lifetime Learning, 1982.
24. Gustafsson L, Hallander HO, Olin P, Reizenstein E, Storsaeter J. A controlled trial of a two-component acellular, a five-component acellular, and a whole-cell pertussis vaccine. *N Engl J Med* 1996;334:349-55.
25. Trollfors B, Taranger J, Lagergård T, et al. A placebo-controlled trial of a pertussis-toxoid vaccine. *N Engl J Med* 1995;333:1045-50.
26. Schmitt H-J, Wirsing von König C-H, Neiss A, et al. Efficacy of acellular pertussis vaccine in early childhood after household exposure. *JAMA* 1996;275:37-41.
27. Edwards KM. Acellular pertussis vaccines — a solution to the pertussis problem? *J Infect Dis* 1993;168:15-20.
28. Liese JG, Harzer E, Meschievitz CK, et al. Case-control study to evaluate the efficacy of BIKEN acellular pertussis vaccine (Tripedia) combined with diphtheria and tetanus toxoids (DTacP) in infants. In: Program and abstracts of the 35th Interscience Conference on Antimicrobial Agents and Chemotherapy, San Francisco, September 17–20, 1995. Washington, D.C.: American Society for Microbiology, 1995:173. abstract.

29. Hallander HO, Storsaeter J, Möllby R. Evaluation of serology and nasopharyngeal cultures for diagnosis of pertussis in a vaccine efficacy trial. *J Infect Dis* 1991;163:1046-54.
30. Edwards KM, Meade BD, Decker MD, et al. Comparison of 13 acellular pertussis vaccines: overview and serologic response. *Pediatrics* 1995;96:548-57.
31. Edwards KM, Decker MD, Halsey NA, et al. Differences in antibody response to whole-cell pertussis vaccines. *Pediatrics* 1991;88:1019-23.
32. Kaplan SL, Lauer BA, Ward MA, et al. Immunogenicity and safety of *Haemophilus influenzae* type b-tetanus protein conjugate vaccine alone or mixed with diphtheria-tetanus-pertussis vaccine in infants. *J Pediatr* 1994;124:323-7.
33. Halperin SA, Bortolussi R, MacLean D, Chisholm N. Persistence of pertussis in an immunized population: results of the Nova Scotia Enhanced Pertussis Surveillance Program. *J Pediatr* 1989;115:686-93.
34. Baker JD, Halperin SA, Edwards K, Miller B, Decker M, Stephens D. Antibody response to *Bordetella pertussis* antigens after immunization with American and Canadian whole-cell vaccines. *J Pediatr* 1992;121:523-7.
35. Fine PEM, Clarkson JA. Reflections on the efficacy of pertussis vaccines. *Rev Infect Dis* 1987;9:866-83.
36. Onorato IM, Wassilak SG, Meade B. Efficacy of whole-cell pertussis vaccine in preschool children in the United States. *JAMA* 1992;267:2745-9.
37. Blennow M, Granström M, Jätmaa E, Olin P. Primary immunization of infants with an acellular pertussis vaccine in a double-blind randomized clinical trial. *Pediatrics* 1988;82:293-9.
38. Cody CL, Baraff LJ, Cherry JD, Marcy SM, Manclark CR. Nature and rates of adverse reactions associated with DTP and DT immunizations in infants and children. *Pediatrics* 1981;68:650-60.
39. Decker MD, Edwards KM, Steinhoff MC, et al. Comparison of 13 acellular pertussis vaccines: adverse reactions. *Pediatrics* 1995;96:557-66.